



A review on the fate, human health and environmental impacts, as well as regulation of antibiotics used in aquaculture

Tijo Cherian^{a,*}, Chinnasamy Ragavendran^b, Smitha Vijayan^a, Sini Kurien^a, Willie J.G. M. Peijnenburg^{c,d,**}

^a School of Biosciences, Mar Athanasios College for Advanced Studies (MACFAST), Tiruvalla, Kerala 689101, India

^b Department of Conservative Dentistry and Endodontics, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences (SIMATS), Chennai 600077, India

^c Institute of Environmental Sciences (CML), Leiden University, Leiden, RA 2300, the Netherlands

^d Center for Safety of Substances and Products, National Institute of Public Health and the Environment (RIVM), P.O. Box 1, Bilthoven, the Netherlands

ARTICLE INFO

Keywords:

Antibiotic
Antibiotic/drug resistance
Environment
Human health

ABSTRACT

Antibiotics have been a necessary component of animal husbandry and aquaculture since they were first used in clinical settings in the 1940s to meet the rising demand for foods generated from animals. Because of this, large-scale industrial animal production has become a hotspot for the evolution and spread of ARGs (antibiotic resistant genes), potentially posing a threat to public health. The recent advent of quick molecular technologies has substantially increased our understanding of ARGs in cattle systems. From a One Health perspective, detailed analyses of ARGs in the livestock industry and potential mitigation strategies are currently available. In order to clarify the intricacies of ARGs across animals, habitats, and people, this review is focused on human health hazards related to antibiotic usages, ARGs in cattle and aquaculture systems. This review specifically addresses the following topics: (1) antimicrobials used in the animal sector; (2) development of ARGs on animals affected by selected agents; (3) ARG transmission mechanisms (direct/indirect animal-to-human); and (4) mitigating strategies. We emphasize the difficulty of reducing the administering antibiotics to animals for the sake of public and environmental health, as well as the critical necessity to take immediate action to stop the transmission of antibiotic/drug resistance in the livestock and aquaculture sectors.

1. Introduction

The term ‘aquaculture’ (latin word ‘aqua’ meaning water) is generally circumscribed as the commercial rearing of aquatic creatures, primarily fish, mollusks, and crustaceans, in water bodies like seas, lakes, ponds, and rivers (FAO 2016b, c; Shao et al., 2021). It traces its historical origin to nearly 4000 years ago, originating in China and stemming out as a propitious economic zone in recent times (Dias et al., 2012), emphasizing strenuous enquiries on on-field practices of research and innovation. According to FAO, an estimated 45% of fishes were produced worldwide via aquaculture practices, with an annual growth rate of > 8% (FAO 2016b, c). The largest producer of aqua-products was China, accounting for about 15% share of the total aquaculture production followed by Indonesia (8%), Peru (7%), Russian Federation (6%), India (5%), USA (5%), Vietnam (4%), Norway (3%), South Korea

(2%), Japan (2%) and Philippines (2%) (FAO 2022) (Fig. 1). Apart from food utilities, the practice of aquaculture has led high economic gains for the traditional fishing activities (FAO 2016a), encouraging the convenient and accessible safe-quality food production for ever-expanding masses, generating jobs and providing a fiscal contribution to overall international economic development. Despite all these pros, a notable ‘call of concern’ need to be addressed pertaining to the matters of environmental impact and risk assessment (Martinez-Porchas and Martinez-Cordova, 2012; Shao et al., 2021). Some of the deleterious effects are listed as: obliteration of natural ecosystems (Rajitha et al., 2007; Martinez-Porchas and Martinez-Cordova, 2012), soil pH changes (Rodriguez-Valencia et al., 2010), water body pollution (Paez-Osuna 2001; Avnimelech and Kochba 2009), chemical contamination (Justino et al., 2016), biological deterioration brought on by the introduction of non-native species (Molnar et al., 2008), landscape variation

* Corresponding author.

** Corresponding author at: Institute of Environmental Sciences (CML), Leiden University, Leiden, RA 2300, the Netherlands.

E-mail addresses: tvarghese891@gmail.com (T. Cherian), peijnenburg@cml.leidenuniv.nl (W.J.G.M. Peijnenburg).

<https://doi.org/10.1016/j.envadv.2023.100411>

Received 23 June 2023; Received in revised form 31 July 2023; Accepted 18 August 2023

Available online 19 August 2023

2666-7657/© 2023 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

(Dumbauld and McCoy 2015), and deteriorating fishery management (Granada et al., 2016). Also, the emergence of new illnesses has coincided with a concomitant increase in the usage of antibiotics in aquaculture productivity practices (Lafferty et al., 2015; Zhang et al., 2023).

The aquatic environment is a dynamic medium capable of long-distance pathogen transfer (Murray, 2013; Chen et al., 2020; Bombaywala et al., 2021), owing to substantial organism movement, and thereby crafting a vastly multifarious web of disease spread (Murray, 2013; Bombaywala et al., 2021). In sequence, for orchestrating the stratagem of ‘control and combat’ of infectious diseases, hefty quantities of veterinary medicinal products (VMPs) (mainly antibiotics) are administered (both controlled and indiscriminate), posing consequential risk to environmental biota and residents of eco-domains (Huang et al., 2015; Lafferty et al., 2015; Xu et al., 2021). Interestingly, many different types of antibiotics have been/are currently being used in aquaculture farms in major producing nations (Fig. 2). Many formulations of antibiotics have been designed and consistently applied as bath treatments or feed augments, devising ‘prevent and treat’ approaches conferring prophylactic and therapeutic defense lines against bacterial infections (Zhang et al., 2023). In the present review, the effect, regulation and fate of antibiotics in environment and their mitigation strategies were explicitly presented using the newly accessible data. The study also points out critical gaps in our current understanding and future areas in need of investigation in the hopes that this section will inspire readers to

get engaged in this hotly debated and rapidly developing field.

2. Methodology: literature search

Screening, identification, and suitability were the three steps in the systematic search approaches used in the present study. The databases of Web of Science, Scopus, and Pubmed/Medline databases, without regard to publication year limits, were used to search for articles specifically about looking into effect of antibiotics in environment. The suitable publications were downloaded in PDF (Portable Document Format) form with conditions of open access or the associated authors were contacted to obtain the full text during the English language literature search. The following were the inclusion requirements: (i) full-text accessibility; (ii) papers that described the methods for preparing, extracting, and analyzing the chosen samples; (iii) papers that described the effects, regulation and entry of antibiotics in environmental domains and that provided information on the mitigation protocols; and (iv) peer-reviewed scientific studies in English language.

3. Entry route into the environment

According to scientific literary estimates, about 75% of feed administered antibiotics enter the environmental domain via (a) metabolic progression of ‘secretion and excretion’ from cultured species, and

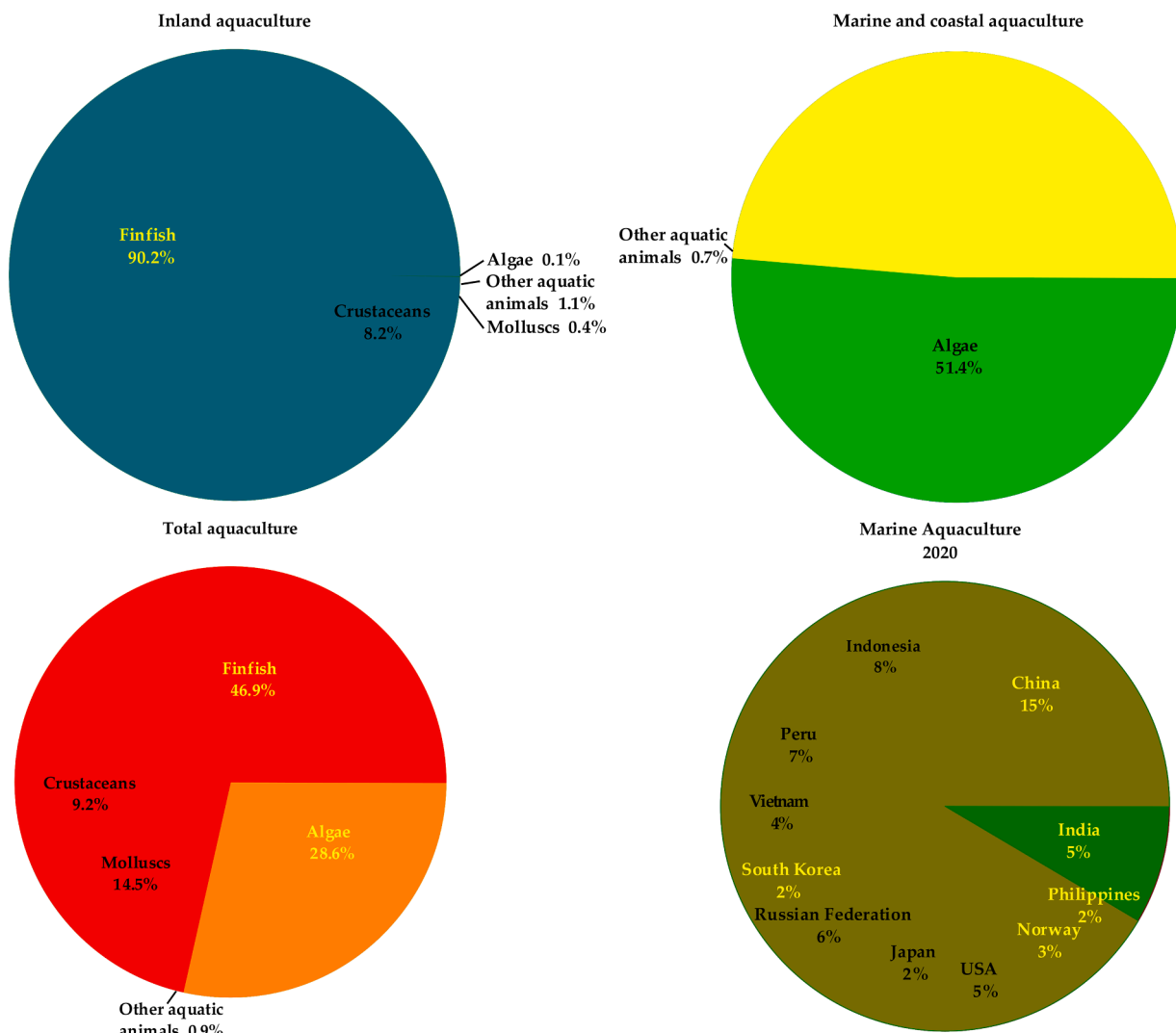


Fig. 1. Aquaculture production data in 2020 (left) and global ten aquaculture-producing countries in 2020 (right) (FAO 2022).

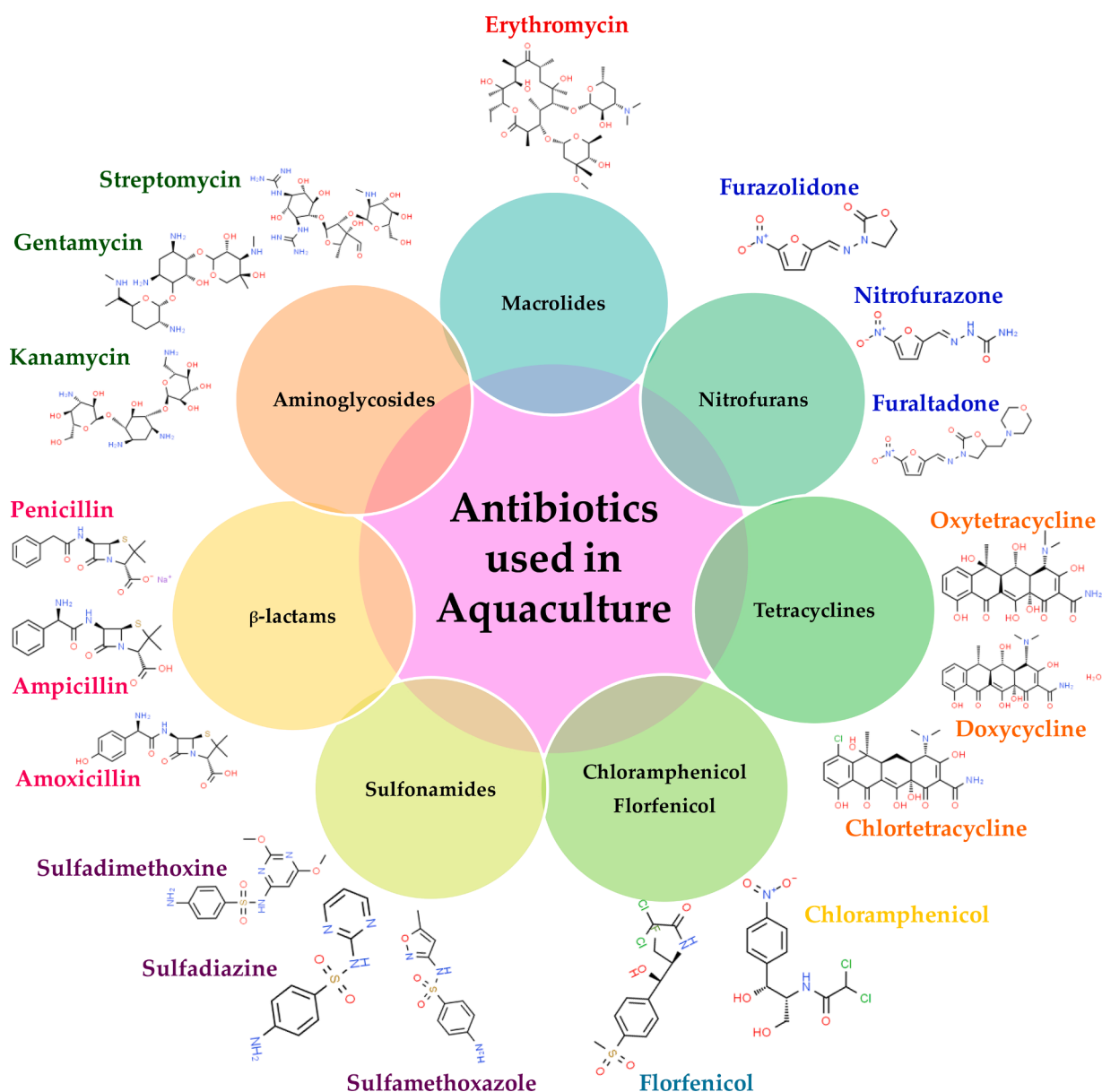


Fig. 2. Antibiotics used in aquaculture.

(b) leaching from unconsumed medicated feed (Lalumera et al., 2004). Antibiotic use and its consequences on aquatic environments have grabbed monumental scientific scrutiny due to their aquaculture-induced pollution and their profound usage in animal husbandry and as human medicine (Santos et al., 2010; Ahmad et al., 2022). The presence of antibiotics has been found to be highly pestilential to microbial entities and primary producers (Zounková et al., 2011). Several toxicological investigations have compiled evidences documenting that aquaculture antibiotics are not hazardous to invertebrates or fish at quantities relevant to the environment (Zounková et al., 2011), although their detrimental effects cannot be eluded on a long-term basis. The study conducted by Wollenberger et al., (2000) discerned for instance lasting effects of oxalinic acid in invertebrates such as disruptive reproduction reported in daphnids at lower concentrations. Fig. 3 depicts entry routes of antibiotics in the environment.

3.1. Entry and fate of antibiotics in soil systems

The effectiveness of antibiotics in the environment is influenced by a range of environmental parameters, including soil type, soil chemistry,

climate, and physico-chemical characteristics (Shao et al., 2021). The fate and behavior of antibiotics in soil have been identified as one of the newer challenges in environmental chemistry (Table 1). Animals administered antibiotics for veterinary purposes excrete the drugs, which then find their way into the soil when the animals graze or when manure is utilized as fertilizer for farming (Chaturvedi et al., 2021; Ahmad et al., 2022; Zhang et al., 2023). Large quantities of antibiotics per hectare have been evaluated to be lost by manuring. Antibiotics in the dust from exhaust air from stable ventilations may be a major source of other minor agricultural environmental releases. Antibiotics are frequently discharged into the environment with very minor transformations, or even completely unaltered and conjugated to polar compounds (Shao et al., 2021; Chaturvedi et al., 2021). The molecular configuration of pharmaceutical antibiotics affects the chemical and physical behavior in soil. Antibiotics are either ionized, amphoteric or amphiphilic, according to their different structural categories. Several physico-chemical characteristics, such as their molecular arrangement, shape, size, hydrophobicity, and solubility differentially affect the binding, fixation and sorption capacity of antibiotics in soils (Shao et al., 2021). Many antibiotics are polar, only weakly water soluble, and

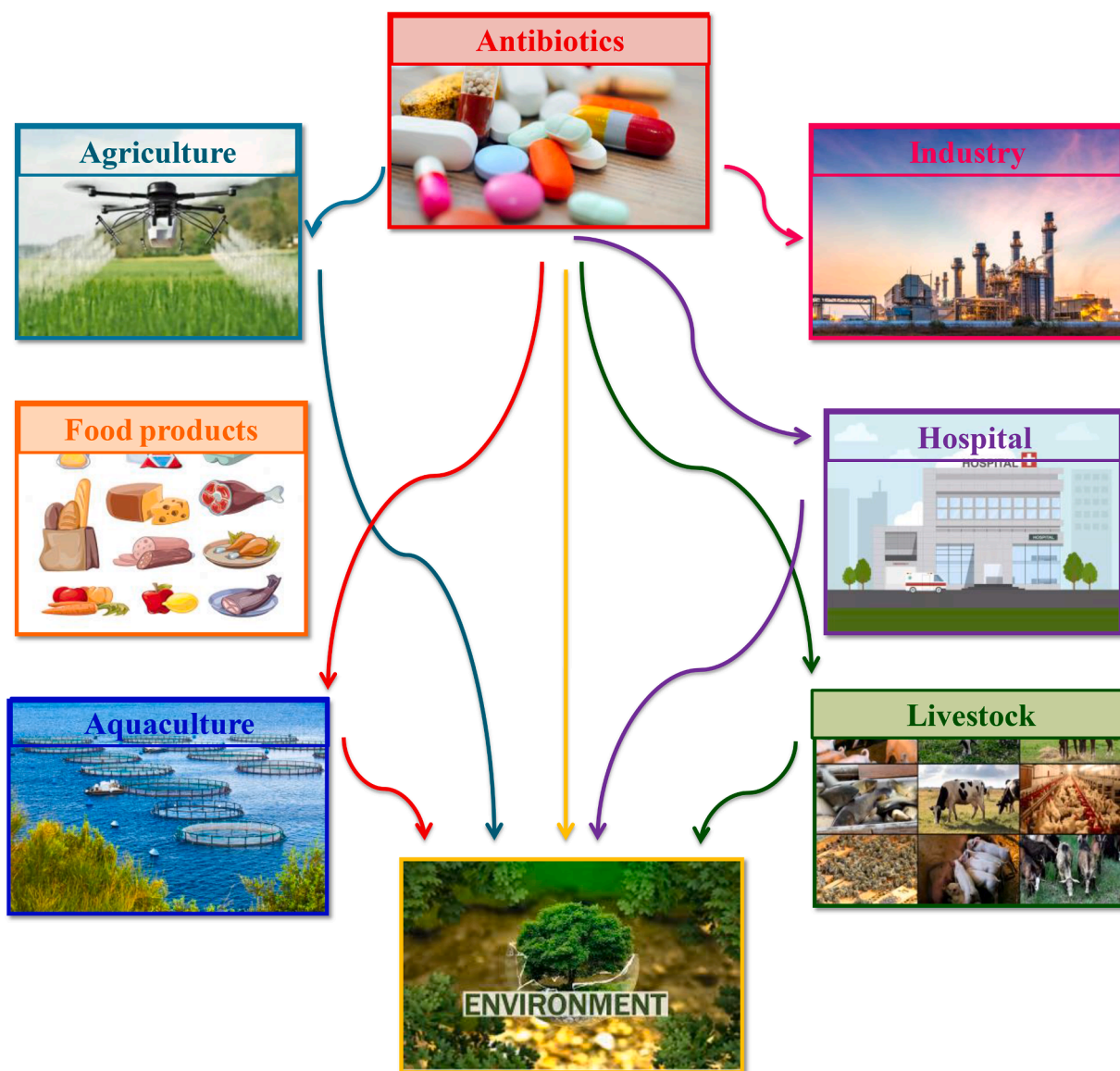


Fig. 3. Entry routes of antibiotics in the environment.

Table 1
Antibiotics in soil systems.

| Class | Antibiotics | Refs. |
|------------------|-------------------|---|
| Macrolides | Clarithromycin | Schüller, 1998 |
| | Lincomycin | Boxall et al., 2004 |
| Sulphonamides | Sulphadiazine | Boxall et al., 2004 |
| | Sulphadimidine | Höper et al., 2002 |
| | Sulphametazine | Hamscher et al., 2005 |
| Trimethoprim | | Boxall et al., 2004 |
| Fluorochinolones | Ciprofloxacin | Schüller, 1998 |
| Tetracyclines | Tetracycline | Winckler and Grafe, 2000; Hamscher et al., 2002 |
| | Oxytetracycline | Boxall et al., 2004 |
| | Chlortetracycline | Hamscher et al., 2005 |

exhibits significant retardation in soils. For instance, the transport of tetracyclines seems to be constrained to quick preferential macropore flow or aided by co-transport by itinerant colloids like dissolved organic

materials (Xu et al., 2021). The majority of antibiotics are quickly absorbed and their efficacy is largely diminished by fixation and sorption, although that does not necessarily entail a full eradication of the antibiotics activity (Zhang et al., 2023). Further, research on the antimicrobial activity of tylosin and tetracycline bound to soil has revealed that these antibiotics are still functionally active even when tightly adsorbed to clay particles (Chaturvedi et al., 2021; Xu et al., 2021). This research demonstrated the antimicrobial impacts that lead to the emergence of antibiotic resistant bacteria in the terrestrial environments. In general a chemical compound usually accumulates when its application rate exceeds its removal rate. The amount of antibiotics found in particular soil layers is referred to as ‘terraccumulation’. These sorbed substances provide a source of contaminants that can be released into soils and affect ground water via leaching or leaching into surface waters (Chen et al., 2020; Bombaywala et al., 2021).

Until now, only a few studies have been reported on the movement and transport of antibiotics in soil. Paez-Osuna (2001) reported the antibiotic contamination of surface waters by diffusion and leakage from agricultural soils. Analysis of ground water and pastoral leachates with manuring and intensive livestock production identified nil to only a few antibiotics in lower concentrations (Bombaywala et al., 2021). In case of

tylosin and oxytetracycline, the distributional coefficient was found to be less manure as compared to the soils (Xu et al., 2021). Similar report was also found in sulphachloropyridazine where likewise a lowering effect of manure was ascertained. An increase in the amount of manure in the soil decreases the distribution coefficient, primarily because of the basic nature of manure. Tetracycline was detected for long time periods up to 30 cm soil depth demonstrating its persistence and accumulation in the environmental system (Xu et al., 2021).

Many factors contribute to the degradation of antibiotics. Since the influence of light is diminished when antibiotics are shielded in slurry or sludge, the process of photodegradation does not yield significant effects (Li et al., 2021). Soil based degradation is primarily sponsored by microbe mediated enzymatic reactions, transforming the native compound via the processes of oxidative decarboxylation and hydroxylation. Although these responses are reversible in nature, the antibiotics subsequently degrade in soil and manure (Zhang et al., 2023). The process of biodegradation increases when manure or sludge containing high microbial load is added to the soil. Soil serves as the storehouse of micro-organisms. High bacterial populations are crucial for maintaining the processes of mineral immobilization and breakdown. Antibiotics in soil usually affects in two ways: first, the microbial ecology may be significantly disrupted; second, these environmental bacteria may acquire up and offer gene-coding resistant factors. Certain soil microorganisms have a built-in resistance to antibiotics (Chen et al., 2020; Shao et al., 2021).

3.2. Entry and fate of antibiotics in water systems

The need for reliable and secure drinking water is growing due to the demand for the planet Earth's limited freshwater resources, especially in view of the continued exponential development of the human population (Xu et al., 2021). The presence and destiny of antibiotics in the water environments have recently been the focus of numerous studies conducted around the globe (Table 2). In the context of antibiotics used in livestock husbandry, the metabolites or degradation products of antibiotics enter the water cycle through the application of manure or slurry to farming used areas, or directly through the excretion of pasture-raised animals on the land, which is then followed by driftage, surface run-off, or leaching into the deeper earth (Caban and Stepnowski, 2021). The aquatic environment may become contaminated with antibiotics from soils. Since the majority of antibiotics are water-soluble, 90% can be eliminated in urine and 75% through animal faeces. Aquaculture is another method by which antimicrobial substances used on animals might be discharged into the environment. Aquaculture residuals and resistant bacteria have been reported in recent years (Okeke et al., 2022). However, the effects of sub-inhibitory amounts against non-marine aquatic bacteria are generally unknown. The influence of several antibiotics continuing to be active against bacteria living in wastewater has been described. The use of sewage sludge as wastewater or manure in irrigation may have directly introduced resistant and multi-resistant bacteria into the food chain. Resistant and multi-resistant bacteria have been detected in wastewater and sewage treatment plants. Most of the investigated antibiotic compounds have been persistent under test settings in aquatic systems, whereas only a small number of antibiotics are partially degraded (Zhang et al., 2023).

Due to the ease of precipitation and accumulation of tetracyclines and hydrolysis of penicillin, they are not present in aquatic environments (Okeke et al., 2022). The β -lactam ring, which is a structural component of β -lactams like cloxacillin, benzylpenicillin, and penicillin, makes them unstable in the environment because β -lactamase, a common enzyme in bacteria, hydrolyzes and breaks open the ring structure (Ahmad et al., 2022). As a result, pristine penicillin molecules are typically absent from the environment. However, antibiotics in agriculture serve as the smallest source of antimicrobials in the marine environment. Only a small number of samples are supposed to have an effect upon animal husbandry on the prevalence of antibiotics in surface

Table 2
Antibiotics in water systems.

| Class | Antibiotics | Source | Refs. |
|------------------|-------------------|---|---|
| Macrolides | Lincomycin | Surface water | Kolpin et al., 2002; Boxall et al., 2004 |
| | Clarithromycin | Surface water | Hirsch et al., 1999 |
| | Erythromycin | Surface water | Hirsch et al., 1999 |
| | Roxithromycin | Surface water | Hirsch et al., 1999 |
| | Tylosin | Surface water | Daughton and Ternes, 1999; Ashton et al., 2004 |
| Sulphonamides | Sulphadiazine | Surface water | Boxall et al., 2004 |
| | Sulphamethazine | Ground water | Hamscher et al., 2005 |
| | Sulphamethoxazole | Ground water Surface water Drinking water | Sacher et al., 2001 Hirsch et al., 1999 Mückter, 2006 |
| Trimethoprim | | Surface water | Boxall et al., 2004 |
| | | Surface water | Hirsch et al., 1999 |
| Fluoroquinolones | Ciprofloxacin | Effluents | Golet et al., 2001 |
| | Norfloxacin | Effluents Surface water | Golet et al., 2001 Kolpin et al., 2002 |
| Tetracyclines | Tetracycline | Ground water | Krapac et al., 2005 |
| | Oxytetracycline | Overland flow water | Kay et al., 2005 |
| | Chlortetracycline | Surface water | Kolpin et al., 2002 |

waters; the majority of the compounds under investigation came from discharge or sewage into rivers. Human administration of antibiotics via municipal wastewater or hospital effluents accounts for the majority of antibiotic input (Li et al., 2021).

4. Antibiotic use in aquaculture: health and environmental safety issues

Although the use of antibiotics benefits enhanced production and the extension of the aquaculture industry, they also carry the vices of harmful effects and risk on human and environmental health (Rico et al., 2013). Scores of scientific literatures have tendered confirmatory accounts on the antibiotic buildup in farmed animal tissues and accumulation in culture and sediments biota, with impending perilous consequences for environmental and human health (Heuer et al., 2009; Shimizu et al., 2013).

4.1. Effect of antibiotics on the environment

The rate of metabolic absorption of orally feed administered antibiotics is low to average (Rico et al., 2013; Wang et al., 2015) and nearly 30–90% of the antibiotics gets excreted via the feces or urine (Sarmah et al., 2006). Besides that discarded food seeps into the environment and contains metabolites of antibiotics, fish faeces, and other degradation products. In intensive fish farm practices, over 70% of the antimicrobials amended to feed diffuse into the environment (Thuy and Nguyen 2013; Andrieu et al., 2015; Giang et al., 2015). In the aqueous environment, the antibiotics and their residual products accumulate in sediments, driving significant changes in the native microbial communities and

perturbing the existing established natural equilibrium (Samuelsen et al., 2014). Conversely, the continual persistence of multi-drug resistance facades in severe environmental implications to overall microbial profiling and ecological stability (Tamminen et al., 2011).

Once introduced in aquatic ecosystems, the antibiotics entail grave and noxious effects on non-target/indicator species particularly, the diverse populations of microalgae, phytoplanktons and zooplanktons (Yasser and Adli 2015; Song et al., 2016). They disrupt early developmental stages of zooplanktons (Park and Kwak 2018) and chlorophyll production in phytoplanktons (Song et al., 2016), convulsing existing egalitarian complexes of food chains and food webs and deranging every level of ecosystem matrices (Cabello 2004). Also, they have been implicated with growth impairment, immunosuppression and altered gut microbiota in farm cultured species (He et al., 2012a).

Algae are oxygenic autotrophs serving as components that are crucial to aquatic habitats and create organic materials that form the bulk feed for most other life forms like fishes and invertebrates. The structural and functional dynamics of an ecosystem can be adversely affected by any biological or chemical changes to algal cells, leading to oxygen depletion and decreased primary output (Ma et al., 2006). One of the most often used ecotoxicological assays for toxicity evaluation in connection to chemical classification and risk assessment is the algal growth inhibition test (EC, 2003; OECD, 2011). Several studies have typified the concentration ranges which impact microalgal growth to be around one or two orders below those toxic concentration ranges for invertebrates (Lai et al., 2009). Among the diversified complex phyto-planktonic communities, cyanobacteria have been found to be the most susceptible group (Brain et al., 2008), due to their speculative morphological resemblance to the target microorganisms. The drug contamination usually affects all living organisms as algal cells forms the base of the food supply and any deficit in their production will have an immediate impact on the entire aquatic food chain. Further, studies pertaining to antibiotic pollution also pinpoint the negative effects on water-quality parameters and the configuration of innate bacterial communities, severely plaguing the functional processes like nitrification and mineralization of organic matter (Tello et al., 2010). Moreover, the relentless and overuse of antibiotics has forayed in a more gruesome theme of antimicrobial resistance in the strains of bacteria, heftily compromising the efficacy of curative treatments and public health. Tendencia and De la Peña (2001) and Le and Munekage (2004) reported that shrimp farms of Philippines and Vietnam had microorganisms that were resistant to antibiotics. Also, the rampant and pervasive use of bulk quantities of antibiotics have resulted in their high bioaccumulation rate via food chains, which led to derived secondary effects in the succeeding apex orders (Cabello, 2006).

4.2. Effects of antibiotics on human health

An unchecked usage of antibiotics in farmed aquaculture-based food production has shown to be a potential hazard for food safety (Chen et al., 2015, 2018b), reducing the scale of marketability and fiscal gains of aquaculture products (Hassan et al., 2013). Additionally, as wild fish can consume residues of antibiotics, the safety of products from catch fisheries is jeopardized (Chiesa et al., 2018). The uncontrolled use and consumption of antibiotic residue laced aquaculture products accounts for an advanced pattern of antibiotic resistance in clinical pathogens and ADRs (adverse drug reactions) (Liu et al., 2017). Li (2008) documented antigenicity of antibiotics like Penicillin G, tetracycline and sulphonomides rooting severe allergies to consumers; while their bioaccumulation induced organ lesions, resulting in chronic toxicity (Zheng and Su 2010). Also, the transcripts concerning occupational health hazards have long term inhalation or close contact of dust aerosols containing antibiotics by manpower in feed mills and cage farms in aquaculture facilities, causing allergies and toxicity and altering normal dermal flora by ingestion and skin contact (White and McDermott 2009). Furthermore, the findings of Moreau and Neis (2009) and Phu

et al., (2016) categorized antimicrobial-occupational hazards as (a) organ-specific and (b) systemic reactions (either singly/ in combinations). Another health hazard referred to as antimicrobial resistance with potential for zoonosis (Martinez 2009), comes typically from long-term high-dose therapy, which causes tissue residue accumulation (Chuah et al., 2016; Monteiro et al., 2016). The infection of *M. albus* by *Aeromonas hydrophila*, followed by red sore disease in *C. carpio* (Saitanu and Wongsawang, 1982), and ulcer disease in walking catfish (*Clarias batrachus*) (Saitanu and Chalarak 1983), were classified as the early imprints of antibiotic resistance by Reungprach and Kesomchandra (1983) from Thailand.

The antimicrobial resistance has been detected in aquaculture across the world in recent research, which is compelling evidence that this danger to human and environmental health is growing.

The culture fish ponds are conceived as the reservoirs of ARGs (antibiotic resistance genes) (Table 3), as the majority of farm used antibiotics (usage either within or outside aquaculture facilities) have been implicated in the development of antimicrobial-resistant microorganisms (Hong et al., 2018; Millanao et al., 2018). In addition, compared to bacteria in areas where there are no aquaculture activity, resident bacteria in commercial aquaculture facilities showed higher ARGs (Gao et al., 2012). The rationale is as follows: Long-term exposure to low doses of antibiotics is necessary for the spread of antibiotic resistance to human biological systems; this is suggestive of the potential for resistance in intestinal resident bacteria and suggests that ARGs may spread horizontally by being conjugated with human pathogenic strains (FAO 2005; Tomova et al., 2015). Moreover, Marshall and Levy (2011) and Rico et al. (2013) exemplified direct transmission of resistant bacterial strains from animals to humans. He et al., (2015) reported the presence of bacteria resistant to antibiotics in aquaculture products in seafood markets of China, hoisting up a major word of consternation in context of health (both of consumers and aquaculture farm workers).

5. The enigmatic problem of multi-antimicrobial resistance (MAMR)

The development of multi-antimicrobial resistant (MAMR; Fig. 4) bacterial strains has severely hampered the existing remedial strategies (Chuah et al., 2016; Pham et al., 2018). The malefic pathogenic strains appropriate resistance to existing schemes of antibiotics, consequential in hysterical breakouts of epidemics and epizootics which are practically terminal and untreatable (Hawkey, 2008). Due to the blanket and wild usage of drug formulations, the MAMR pattern has been confirmed for two *Aeromonas caviae* HG4 isolates (Rahman et al., 2009); members of Bacilli (Fernandez-Alarcon et al., 2010), *V. parahaemolyticus* and

Table 3
Country-specific number of studies on antibiotic resistance among top ten nations.

| Nation | No. of studies | % | Refs. |
|-------------|----------------|------|---|
| China | 74.0 | 43.5 | Liang et al., 2013; Su et al., 2017; Chen et al., 2018a; Hong et al., 2018; Marti et al., 2018 |
| Indonesia | 1.0 | 0.6 | Zulkifli et al., 2009 |
| India | 20.0 | 11.8 | Elmahdi et al., 2016; Rahiman et al., 2016; Sneha et al., 2016; Stratev and Odeyemi 2016; Mishra et al., 2017 |
| Vietnam | 21.0 | 12.4 | Son et al., 2011; Nguyen et al., 2014, 2016; Pham et al., 2018; Thai et al., 2018 |
| Philippines | 1.0 | 0.6 | Elmahdi et al., 2016 |
| Bangladesh | 2.0 | 1.2 | Rahman et al., 2009; Hossain et al., 2017 |
| South Korea | 5.0 | 2.9 | Yi et al., 2014; Germond and Kim 2015; Jang et al., 2018; Kim et al., 2018 |
| Egypt | 4.0 | 2.4 | Ishida et al., 2010; Ali et al., 2016; Osman et al., 2016, 2017 |
| Norway | 2.0 | 1.2 | Burrige et al., 2010; Midtlyng et al., 2011 |
| Japan | 3.0 | 1.8 | Sun et al., 2010; Ahmed et al., 2015; Cao et al., 2016 |

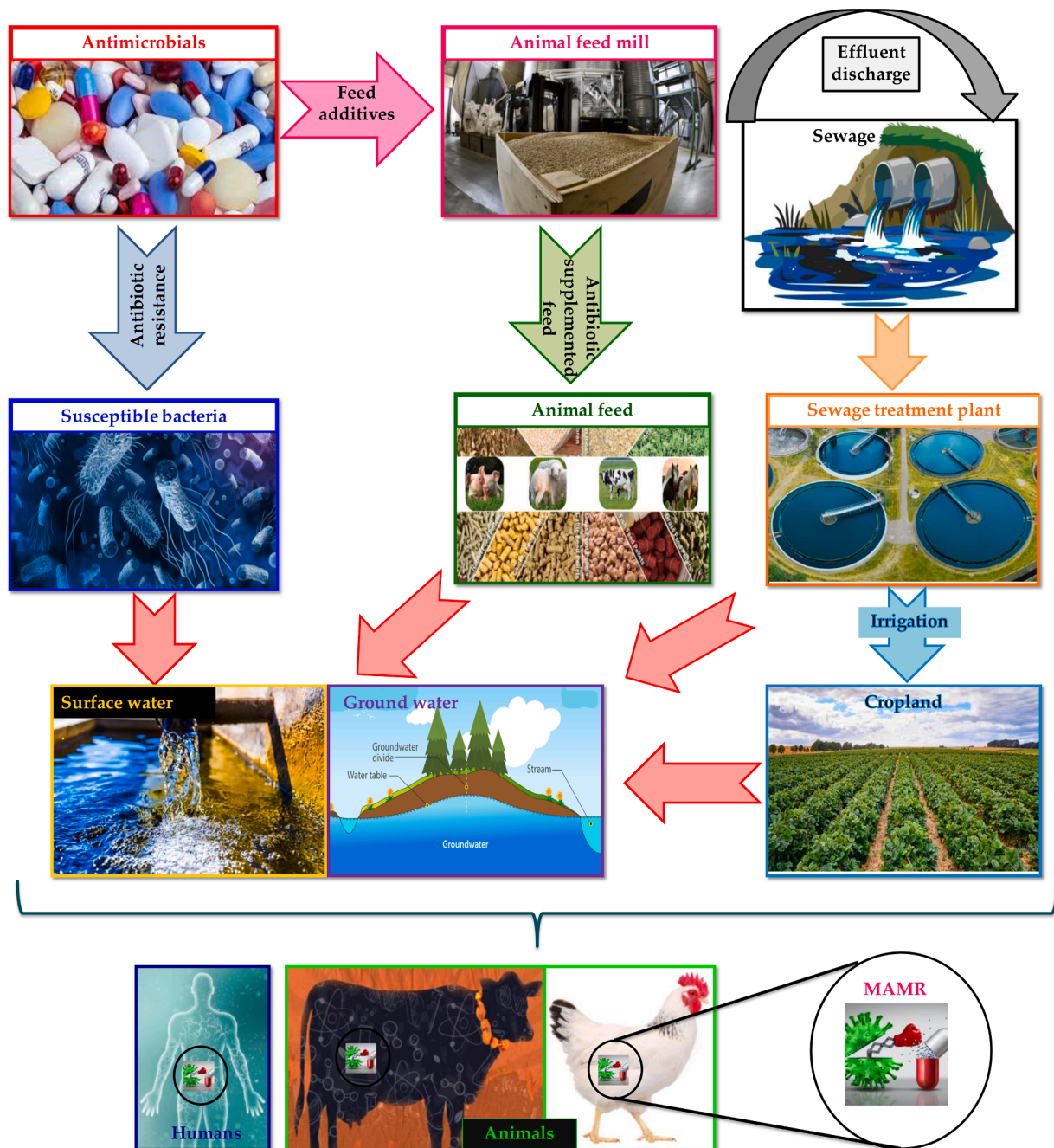


Fig. 4. Environment-to-animal-to-human MAMR transmission pathways in the One Health framework.

V. cholerae (Noorlis et al., 2011); *Vibrio* isolates (Rocha et al., 2016); *V. vulnificus* and *V. parahaemolyticus* (Elmahdi et al., 2016); *Klebsiella pneumoniae* and *A. hydrophila* (Pham et al., 2018). Also, the scenario of MAMR has been reported from coastal sea waters and estuaries for the species of *Vibrio* sp. (*V. parahaemolyticus* and *V. alginolyticus*), *Acinetobacter*, *Photobacterium* sp., *Aeromonas* and *Klebsiella* (Germond and Kim 2015; de Menezes et al., 2017).

It has been widely noted that plasmid mediation and integron transfer are the two ways by which antibiotic resistance spreads across bacteria (Table 4). The scientific confirmation of integron-mediated MAMR genes has been substantiated in the members of the family of Enterobacteriaceae, *E. coli* (Deng et al., 2014, 2016), *E. coli* isolates from China (Chen et al., 2011), and *Aeromonas* spp. Alternatively, the plasmid-mediated MAMR transfer was verified in the isolates of *Vibrio* (Rocha et al., 2016); *Salmonella* (Budiati et al., 2013) and

V. parahaemolyticus (Letchumanan et al., 2015a, b). Plasmid mediated single antibiotic resistance was reported in the species of *Pseudomonas*, *Salmonella* serovars (Budiati et al., 2013), *Edwardsiella ictaluri* (Dung et al., 2008, 2009), *Aeromonas* spp. (Nguyen et al., 2014) and *Flavobacterium psychrophilum* (Henríquez-Nuñez et al., 2012). The presence of antibiotic-resistant *Escherichia coli* (Shah et al., 2014), *Aeromonas*, *Acinetobacter* (Agersø et al., 2007), *Vibrio* (Reboucas et al., 2011; Tuševljak et al., 2013), isolates of *Streptococcus* and *Enterococcus* (Osman et al., 2017), *Salmonella* and *Edwardsiella* (Tuševljak et al., 2013) have also been reported from aquaculture farmed organisms. Above all, the unbridled prophylactic usage of antibiotics/drug formulations in aquaculture elevates resistance menace due to amplified selective pressure, rendering the progressive ineffectiveness of drugs (Schwarz et al., 2006). Also, due to the inherent and intrinsic unions of aquaculture systems with open water bodies, the antibiotic-resistant

Table 4
Prevalence of multi-antimicrobial resistance (MAMR).

| Mode of resistance | MAMR isolates | Refs. |
|---|-------------------------------------|---|
| Plasmid mediation | <i>Aeromonas caviae</i> HG4 | Rahman et al., 2009 |
| Plasmid mediation | Bacilli isolates | Fernandez-Alarcon et al., 2010 |
| Plasmid mediation | <i>V. parahaemolyticus</i> | Noorlis et al., 2011 |
| Plasmid mediation | <i>V. cholerae</i> | Noorlis et al., 2011 |
| Plasmid mediation | <i>Vibrio</i> isolates | Rocha et al., 2016 |
| Plasmid mediation | <i>V. vulnificus</i> | Elmahdi et al., 2016 |
| Plasmid mediation | <i>V. parahaemolyticus</i> | Elmahdi et al., 2016 |
| Plasmid mediation | <i>Klebsiella pneumoniae</i> | Pham et al., 2018 |
| Plasmid mediation | <i>Aeromonas hydrophila</i> | Pham et al., 2018 |
| Plasmid mediation | <i>Vibrio parahaemolyticus</i> | Germond and Kim 2015 |
| Plasmid mediation | <i>Vibrio alginolyticus</i> | Germond and Kim 2015 |
| Plasmid mediation | <i>Acinetobacter</i> sp. | Germond and Kim 2015 |
| Plasmid mediation | <i>Photobacterium</i> sp. | de Menezes et al., 2017 |
| Plasmid mediation | <i>Aeromonas</i> sp. | de Menezes et al., 2017 |
| Plasmid mediation | <i>Klebsiella</i> sp. | de Menezes et al., 2017 |
| Integron-mediated MAMR genes | <i>E. coli</i> | Deng et al., 2014, 2016 |
| Integron-mediated MAMR genes | <i>E. coli</i> isolated from China | Chen et al., 2011 |
| Integron-mediated MAMR genes | <i>Aeromonas</i> sp. | Chen et al., 2011 |
| Plasmid-mediated MAMR transfer | isolates of <i>Vibrio</i> | Rocha et al., 2016 |
| Plasmid-mediated MAMR transfer | isolates of <i>Salmonella</i> | Budiati et al., 2013 |
| Plasmid-mediated MAMR transfer | <i>Vibrio parahaemolyticus</i> | Letchumanan et al., 2015a,b |
| Plasmid mediated single antibiotic resistance | <i>Pseudomonas</i> | Budiati et al., 2013 |
| Plasmid mediated single antibiotic resistance | <i>Salmonella</i> serovars | Budiati et al., 2013 |
| Plasmid mediated single antibiotic resistance | <i>Edwardsiella ictaluri</i> | Dung et al., 2008, 2009 |
| Plasmid mediated single antibiotic resistance | <i>Aeromonas</i> sp. | Nguyen et al., 2014 |
| Plasmid mediated single antibiotic resistance | <i>Flavobacterium psychrophilum</i> | Henríquez-Nuñez et al., 2012 |
| Plasmid mediation | <i>Escherichia coli</i> | Shah et al., 2014 |
| Plasmid mediation | <i>Aeromonas</i> sp. | Agersø et al., 2007 |
| Plasmid mediation | <i>Acinetobacter</i> sp. | Agersø et al., 2007 |
| Plasmid mediation | <i>Vibrio</i> sp. | Reboucas et al., 2011; Tuševljak et al., 2013 |
| Plasmid mediation | isolates of <i>Streptococcus</i> | Osman et al., 2017 |
| Plasmid mediation | isolates of <i>Enterococcus</i> | Osman et al., 2017 |
| Plasmid mediation | isolates of <i>Salmonella</i> | Tuševljak et al., 2013 |
| Plasmid mediation | isolates of <i>Edwardsiella</i> | Tuševljak et al., 2013 |

bacterial strains have been well reported from open water systems (Zhang et al., 2014).

6. Mitigation strategies

6.1. Management of certain agents

The most effective way to stop the spread of MAMRs derived from animal use is to minimize and reduce antibiotic administration in the livestock economy. It has been demonstrated that limiting the use of antibiotics and using other corrective measures can significantly slow the spread of MAMRs in cattle production (Tang et al., 2017). Meanwhile, enhancing animal health through improved livestock management, improved farm hygiene, and the use of a preventative vaccine to fend off animal diseases can aid in reducing the need for antibiotics. In order to decrease the consumption of antibiotics and the emergence of antibiotic resistance in animals, additional additives such as prebiotics, probiotics, and phyto-based products may emerge as reliable alternatives (Yi et al., 2020). Also, due to their potent antibacterial properties and minimal capacity to induce resistance, antimicrobial peptides are suggested as a potent replacement for antibiotics (Li et al., 2018). By taking stock with the cattle waste, it is also possible to lessen the release

of MAMRs and antibiotics into the environment. MAMR transmission and proliferation could be less likely in certain environmental niches where antibiotic-driven selection was found to be reduced.

6.2. Management of manure

Fresh manure borne pathogens can be removed through composting, which can also lessen odour emissions and create organic matter for crop productivity. It has been demonstrated that hyperthermophilic composting, which may reach temperatures of up to 90 °C during the fermentation phase, removes MAMRs more effectively than traditional composting (Liao et al., 2018). This may be because of the high temperature reached during the hyperthermophilic phase. Composting can effectively remove antibiotics like penicillin, ionophores, macrolides, fluoroquinolones, sulfonamides, and tetracyclines, thereby reducing the potential selection for MAMRs along the pathways (Selvam and Wong 2017). But nevertheless, due to the intricate microbial ecological processes involved in composting, the behavior of MAMRs changes during this process.

To increase the removal effectiveness of MAMRs, additives including coal gasification slags superabsorbent polymers, zeolites, and biochar can be used in composting or soils (Peng et al., 2018). In particular, biochar can immobilize heavy metals, speedup the breakdown of organic waste, and limits nutrient loss and elevates the thermophilic composting temperature (Li Duan et al., 2017). A number of studies have demonstrated that some prominent characteristics of biochar such as porous structure, high surface-to-volume ratio, and microbial affinity enable it to restrict the multiplication of MAMR in soils and plants (Zhang et al., 2019). It has been demonstrated that the removal of MAMRs using the regularly utilized livestock waste treatment method of anaerobic digestion is both affordable and effective (Couch et al., 2019). During anaerobic digestion, certain resistance genes, such as *tet* and *erm* genes, have reported reductions in their log copy numbers (Couch et al., 2019). However, a number of variables, including substrate types, operational temperatures, and the microbial hosts of MAMRs, have a significant impact on how effectively anaerobic digestion reduces MAMRs in animal waste. Like composting, additives like wheat straw and biochar can enhance the elimination of MAMR during anaerobic digestion by preventing the re-absorption of metals and antibiotic residues (Yi et al., 2020). Despite the fact that composting and anaerobic digestion are important techniques for effectively disposing of MAMRs, it should be noted that they do not completely eliminate MAMRs. In fact, some MAMRs can still surpass treatment process, even with an increase in the prevalence of resistance. In order to prevent the potential MAMRs surfeit, additional procedures or discharge management may need to be taken into account.

6.3. Treatment of waste water

The abundance of MAMRs can be greatly reduced and their spread from animal husbandry, especially aquaculture, can be effectively controlled by effluent treatments from the pastoral farms. MAMR levels in effluents following wastewater treatment reactors have been found to be decreased in the vast majority of studies. A study by An et al. (2018) reported a reduction in the number of resistance genes and integrons cassettes in the wastewater treatment processes. Although wastewater treatment has been found to be a viable approach for eliminating ARGs, developing nations frequently lack adequate wastewater management or treatment. Therefore, the contaminated effluent may be discarded straight to nearby aquatic bodies (Gros et al., 2019). Moreover, the treatment method can have a significant impact on the elimination of MAMRs. For example, animal effluent from ordinary farms is treated in a bioreactor through an artificial wetland without a full-scale wastewater treatment system. In contrast, ARGs are more effectively eliminated in water treatment plants. The quantity of intake, the kind of biological therapies, and the length of the hydraulic residency are all related to the

success of the treatment, influencing the elimination of bacteria and MAMRs (Novo and Manaia, 2010).

6.4. Policies for management and assistance

To manage AMR issues under One Health, regulatory standards for livestock farming and their waste management are crucial. In order to address the worldwide problem of MAMR, it is crucial to involve decision-makers, researchers, farm workers, animal agricultural stockholders, and the community at large in all nations (Góchez et al., 2019; Yi et al., 2020). The discovery of solutions and the provision of a better proposal for concerned authorities on the MAMR danger could be facilitated by standardized procedures and reliable libraries for antibiotics and MAMRs in livestock sectors. Implementing antibiotic limitations may move more quickly if current legislations are improved. This will result in lower financial losses for the livestock sector. Approaches that are both economically viable and environmentally sustainable must be further developed, enhanced, and integrated (Yi et al., 2020). Monitoring and risk evaluations of MAMRs in wastewater and animal waste are required before further applications in order to prevent the spread of MAMRs from animal husbandry to downstream habitats and humans. To combat antibiotic resistance, both regional and global cooperation is required. This includes reiterating the danger that antibiotic resistance poses and searching for new cutting-edge technology or efficient management solutions (Yi et al., 2020). All stakeholders and the general public must be educated and made aware of MAMRs in order to raise awareness and change the current scenario.

Although many decades have passed since the first-time use of antibiotics in feedlots, scientific research on the providence and incidence of antibiotics has only recently emerged from its infancy. Assessments of antibiotic concentrations in the environment and known effects on terrestrial or aquatic microorganisms have not been evaluated, in part due to the lack of appropriate analytical test methods (Yi et al., 2020). The effects of antimicrobial medications in aquatic and terrestrial environments depend not only on the dosage and mode of administration but also on the methods used for animal husbandry, the internal metabolism of the animals, the handling and storage of their manure, and the rates of degradation of the antimicrobials (Gros et al., 2019). Unfortunately, there is a dearth of information regarding the distribution and destiny of antibiotics in the environment, as well as their prevalence.

The propagation of multi-drug resistant bacteria, important for human and veterinary medicine, is one of the greatest issues of prolonged use of antibiotics and their release into the environment (Couch et al., 2019). Pathogenic bacteria may be able to obtain resistance genes from commensal and environmental bacteria. Pathogens, commensal bacteria, and environmental bacteria may develop single, cross, and even multiple resistances as a result of environmental antimicrobials. To find out the role of antimicrobials in the establishment and maintenance of single or multiple populations of antibiotic-resistant bacteria, particularly pathogenic bacteria, further study is still required (An et al., 2018). There is also a lack of reliable information on the relationship between antibiotic residues and the prevalence of resistant bacteria. Even if a general correlation between antibiotic usage and the percentage of resistant strains is anticipated, it is unclear at what threshold concentrations a change towards an increase in resistance bacteria is to be expected. These resistant bacterial strains may spread through direct contact or the food chain, which might lessen the therapeutic effects of using antibiotics to treat both animals and humans (Peng et al., 2018). Emphasis must be placed on multidrug-resistant bacteria's new and existing exposure pathways from animals to humans.

By continuously manuring contaminated faeces from agriculture, it is discovered that resistance is created by repeatedly exposing bacteria to sub-lethal concentrations of antibiotics (Yi et al., 2020). Long-term spreading of liquid manure on fields could lead to major contamination, especially if the environment accumulates specific antibiotics like tetracycline. Despite the fact that certain antibiotics are administered to

both humans and animals, human usage is the primary cause of most human resistance problems (Yi et al., 2020). Livestock animal husbandry pollutes the environment with antibiotic residues and resistant microorganisms, yet it appears that people, rather than animals, are the major source of these poisons. Wastewater treatment facilities are the primary source of antibiotic discharge into the environment since the clearance during treatment is inadequate. According to new strategies, sources of faeces pollution of human or animal origin can be identified by analyzing bacteria for signs of resistance to veterinary or human medications (Li et al., 2017).

Although it is clear that dealing with antimicrobial chemicals is necessary for an effective animal production, antibiotics should never be used as the replacement of an appropriate hygiene management system. As the WHO states, "Enhanced monitoring of antibiotic use by data acquisition at various levels is a step towards a science-based, successful intervention." Furthermore, the environmental hazards exerted by antibiotics may have been only now starting to be identified by studies (Liao et al., 2018). Environmentally relevant doses for the vast majority of chemicals are currently found to be substantially lesser than the real concentrations utilized on target species. Due to the difficulties in evaluating controlled comparisons and the variety of natural systems, the advantages of lowering antibiotic resistance are difficult to quantify.

The scientific community is currently debating the best approach to take in order to solve this issue. Experimentation alone cannot determine whether antibiotics have a negative impact on ecosystem health. Laboratory research frequently has limited applicability to the environment due to differences in parameters which includes moisture content, pH, concentration, temperature, and other circumstances such as in experiments on transport, microbial degradability, and the metabolic route of antibiotics (Zhang et al., 2019; Yi et al., 2020). The data collection from pathway mobility and environmental sampling experiments involving significant veterinary pharmaceuticals is one suggested approach to solving this problem; however, the pertinent data have not yet been enough in the analysis of currently used antibiotics (Li et al., 2017). The scientific findings presented in this review illustrate that the occurrence of resistant bacteria and the extent of antibiotic contamination are not constrained by interspecies barriers, geographical differences, or fiscal conditions, even though this deficiency of financial profit worsens the call for extensive actions by the research community. The long-term invisible impacts on numerous species and other interacting effects on living things and the environment as a whole deserve more study, according to scientists. The sensible use of antibiotics as a preventative measure raises questions about the efficacy of both veterinary and human drug therapy (Tables 2–4).

7. Conclusions

The objective of this study was to convey the current level of knowledge about the risks to the environment and public health induced by the excessive use of antibiotics in aquaculture. The unsystematic and random prophylactic uses of antibiotics along with the incidents of MAMRs have put credence on the assessment of health and environmental risks. The major accents spotlighted were: occupational health hazards, food safety issues, altered biodiversity, and antibiotic resistance. Meanwhile, the blanket use of drug formulations/antibiotics remains high in major producer countries. This underscores the call for stern pursuance and enforcement of regulations and policies. Further, meticulous planning, economic investment in research prospects, coordination of policies and regulations, and international collaboration are necessary for attaining sustainable aquaculture production and fine-tuned groundwork in production and marketing. The reduction of antibiotic consumption per unit in the manufacture of environmentally friendly and securely ingestible aquatic food should be the overarching goal. Also, there is an urgent need to survey and ascertain compound specific usage data in aquaculture production.

CRedit authorship contribution statement

Tijo Cherian: Conceptualization, Investigation, Validation, Writing – original draft, Writing – review & editing. **Chinnasamy Ragavendran:** Validation, Writing – review & editing. **Smitha Vijayan:** Writing – review & editing. **Sini Kurien:** Writing – review & editing. **Willie J.G. M. Peijnenburg:** Writing – review & editing.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

Data will be made available on request.

References

- Agersø, Y., Bruun, M.S., Dalsgaard, I., Larsen, J.L., 2007. The tetracycline resistance gene tet(E) is frequently occurring and present on large horizontally transferable plasmids in *Aeromonas* spp. from fish farms. *Aquaculture* 266, 47–52.
- Ahmad, A., Kurniawan, S.B., Sheikh Abdullah, S.R., Ahmad, R.O., Hasan, H.A., 2022. Contaminants of emerging concern (CECs) in aquaculture effluent: Insight into breeding and rearing activities, alarming impacts, regulations, performance of wastewater treatment unit and future approaches. *Chemosphere* 290, 133319.
- Ahmed, A.M., Maruyama, A., Khalifa, H.O., Shimamoto, T., Shimamoto, T., 2015. Seafood as a reservoir of Gram-negative bacteria carrying integrons and antimicrobial resistance genes in Japan. *Biomed. Environ. Sci.* 28, 924–926.
- Ali, S.S., Shaaban, M.T., Abomohra, A., El-Safy, K., 2016. Macroalgal activity against multiple drug resistant *Aeromonas hydrophila*: a novel treatment study towards enhancement of fish growth performance. *Microb. Pathog.* 101, 89–95.
- An, X.L., Chen, Q.L., Zhu, D., Zhu, Y.G., Gillings, M.R., Su, J.Q., 2018. Impact of wastewater treatment on the prevalence of integrons and the genetic diversity of integron cassettes. *Appl. Environ. Microbiol.* 84 (9), e02766. <https://doi.org/10.1128/AEM.02766-17>.
- Andrieu, M., Rico, A., Phu, T.M., Phuong, N.T., Van den Brink, P.J., 2015. Ecological risk assessment of the antibiotic enrofloxacin applied to *Pangasius* catfish farms in the Mekong Delta, Vietnam. *Chemosphere* 119, 407–414.
- Avnimelech, Y., Kochba, M., 2009. Evaluation of nitrogen uptake and excretion by tilapia in bio flocc tanks, using 15N tracing. *Aquaculture* 287, 163–168.
- Bombaywala, S., Mandpe, A., Paliya, S., et al., 2021. Antibiotic resistance in the environment: a critical insight on its occurrence, fate, and eco-toxicity. *Environ. Sci. Pollut. Res.* 28, 24889–24916.
- Boxall, A.B., Fogg, L., Blackwell, P., Blackwell, P., Kay, P., Pemberton, E., et al., 2004. Veterinary medicines in the environment. Cunningham F, Elliott J, Lees P (Eds) *Reviews of Environmental Contamination and Toxicology*. Springer, Heidelberg, Germany, pp. 1–91.
- Brain, R.A., Hanson, M.L., Solomon, K.R., Brooks, B.W., 2008. Aquatic plants exposed to pharmaceuticals: effects and risks. *Rev. Environ. Contam. Toxicol.* 192, 67–115.
- Budiati, T., Rusul, G., Wan-Abdullah, W.N., Arip, Y.M., Ahmad, R., Thong, K.L., 2013. Prevalence, antibiotic resistance and plasmid profiling of *Salmonella* in catfish (*Clarias gariepinus*) and tilapia (*Tilapia mossambica*) obtained from wet markets and ponds in Malaysia. *Aquaculture* 372–375, 127–132.
- Burridge, L., Weis, J.S., Cabello, F., Pizarro, J., Bostick, K., 2010. Chemical use in salmon aquaculture: a review of current practices and possible environmental effects. *Aquaculture* 306, 7–23.
- Caban, M., Stepnowski, P., 2021. How to decrease pharmaceuticals in the environment? A review. *Environ. Chem. Lett.* 19, 3115–3138.
- Cabello, F.C., 2004. Antibiotics and aquaculture in Chile: implications for human and animal health. *Rev. Med. Chil.* 132, 1001–1006.
- Cabello, F.C., 2006. Heavy use of prophylactic antibiotics in aquaculture: a growing problem for human and animal health and for the environment. *Environ. Microbiol.* 8, 1137–1144.
- Cao, H.P., Long, X.W., Lu, L.Q., Yang, X.L., Chen, B.Y., 2016. *Citrobacter freundii*: a causative agent for tail rot disease in freshwater cultured Japanese Eel *Anguilla japonica*. *Israeli J. Aquac. Bamidgah* 68.
- Chaturvedi, P., Shukla, P., Giri, B.S., Chowdhary, P., Chandra, R., Gupta, P., Pandey, A., 2021. Prevalence and hazardous impact of pharmaceutical and personal care products and antibiotics in environment: a review on emerging contaminants. *Environ. Res.* 194, 110664.
- Chen, B., Zheng, W., Yu, Y., Huang, W., Zheng, S., Zhang, Y., et al., 2011. Class 1 integrons, selected virulence genes, and antibiotic resistance in *Escherichia coli* isolates from the Minjiang River, Fujian Province, China. *Appl. Environ. Microbiol.* 77, 148–155.
- Chen, H., Liu, S., Xu, X.R., Liu, S.S., Zhou, G.J., Sun, K.F., et al., 2015. Antibiotics in typical marine aquaculture farms surrounding Hailing Island, South China: occurrence, bioaccumulation and human dietary exposure. *Mar. Pollut. Bull.* 90, 181–187.
- Chen, B.W., Lin, L., Fang, L., Yang, Y., Chen, E.Z., Yuan, K., et al., 2018a. Complex pollution of antibiotic resistance genes due to beta-lactam and aminoglycoside use in aquaculture farming. *Water Res.* 134, 200–208.
- Chen, H., Liu, S., Xu, X.R., Diao, Z.H., Sun, K.F., Hao, Q.W., et al., 2018b. Tissue distribution, bioaccumulation characteristics and health risk of antibiotics in cultured fish from a typical aquaculture area. *J. Hazard. Mater.* 343, 140–148.
- Chen, J., Sun, R., Pan, C., Sun, Y., Mai, B., Li, Q.X., 2020. Antibiotics and food safety in aquaculture. *J. Agric. Food Chem.* 68 (43), 11908–11919.
- Chiesa, L.M., Nobile, M., Malandra, R., Panseri, S., Arioli, F., 2018. Occurrence of antibiotics in mussels and clams from various FAO areas. *Food Chem.* 240, 16–23.
- Chuah, L.O., Effarizah, M., Goni, A.M., Rusul, G., 2016. Antibiotic application and emergence of multiple antibiotic resistance (MAR) in global catfish aquaculture. *Curr. Environ. Health Rep.* 3, 118–127.
- Couch, M., Agga, G.E., Kasumba, J., Parekh, R.R., Loughrin, J.H., Conte, E.D., 2019. Abundances of tetracycline resistance genes and tetracycline antibiotics during anaerobic digestion of swine waste. *J. Environ. Qual.* 48 (1), 171–178. <https://doi.org/10.2134/jeq2018.09.0331>.
- de Menezes, F.G.R., Rodriguez, M.T.T., de Carvalho, F.C.T., Reboucas, R.H., Costa, R.A., de Sousa, O.V., et al., 2017. Pathogenic *Vibrio* species isolated from estuarine environments (Ceara, Brazil) -antimicrobial resistance and virulence potential profiles. *An. Acad. Bras. Cienc.* 89, 1175–1188.
- Deng, Y., Wu, Y., Tan, A., Huang, Y., Jiang, L., Xue, H., et al., 2014. Analysis of antimicrobial resistance genes in *Aeromonas* spp. isolated from cultured freshwater animals in China. *Microb. Drug Resist.* 20, 350–356.
- Deng, Y., Wu, Y., Jiang, L., Tan, A., Zhang, R., Luo, L., 2016. Multi-drug resistance mediated by class 1 integrons in *Aeromonas* isolated from farmed freshwater animals. *Front. Microbiol.* 7, 935.
- Dias, J.D., Simões, N.R., Bonecker, C.C., 2012. Net cages in fish farming: a scientometric analysis. *Acta Limnol. Bras.* 24, 12–17.
- Dumbauld, B., McCoy, L., 2015. Effect of oyster aquaculture on seagrass *Zosteramarina* at the estuarine landscape scale in Willapa Bay, Washington (USA). *Aquacult. Env. Interact.* 7, 29–47.
- Dung, T.T., Haesebrouck, F., Tuan, N.A., Sorgeloos, P., Bael, M., Decostere, A., 2008. Antimicrobial susceptibility pattern of *Edwardsiella ictaluri* isolates from natural outbreaks of bacillary necrosis of *Pangasius nodonhyppophthalmus* in Vietnam. *Microb. Drug Resist.* 14, 311–316.
- Dung, T.T., Haesebrouck, F., Sorgeloos, P., Tuan, N.A., Pasmans, F., Smet, A., et al., 2009. IncK plasmid-mediated tetracycline resistance in *Edwardsiella ictaluri* isolates from diseased freshwater catfish in Vietnam. *Aquaculture* 295, 157–159.
- EC (2003). Technical guidance document on risk assessment in support of commission directive 93/67 on risk assessment for new notified substances, Commission Regulation (EC) no. 1488/94 on risk assessment for existing substances and Directive 98/8/EC of the Parliament and of the Council concerning the placing of biocidal products on the market. European Commission.
- Elmahdi, S., DaSilva, L.V., Parveen, S., 2016. Antibiotic resistance of *Vibrio parahaemolyticus* and *Vibrio vulnificus* in various countries: a review. *Food Microbiol.* 57, 128–134.
- FAO, 2005. Responsible Use of Antibiotics in Aquaculture. Food and Agriculture Organization of the United Nations, Rome, Italy. Serrano PHFAO Fisheries Technical Paper 469.
- FAO, 2016a. State of World Fisheries and Aquaculture 2016 (Spanish). Food & Agriculture Org. S.I., Rome.
- FAO, 2016b. Contributing to Food Security and Nutrition for All, (The State of World Fisheries and Aquaculture). Food and Agriculture Organization of the United Nations, Rome.
- FAO (2016c) El estado mundial de la pesca y la acuicultura (2016). Contribucion a la seguridad alimentaria y la nutricion paratodos, Roma, 224 pp.
- FAO, 2022. The State of World Fisheries and Aquaculture (2022). Towards Blue Transformation. FAO, Rome. <https://doi.org/10.4060/cc0461en>.
- Fernandez-Alarcon, C., Miranda, C.D., Singer, R.S., Lopez, Y., Rojas, R., Bello, H., et al., 2010. Detection of the floR gene in a diversity of florfenicol resistant gram-negative bacilli from freshwater salmon farms in Chile. *Zoonoses Public Health* 57, 181–188.
- Góchez, D., Raicek, M., Pinto Ferreira, J., Jeannin, M., Moulin, G., Erlacher-Vindel, E., 2019. OIE annual report on antimicrobial agents intended for use in animals: methods used. *Front. Vet. Sci.* 6, 317. <https://doi.org/10.3389/fvets.2019.00317>.
- Gao, P., Mao, D., Luo, Y., Wang, L., Xu, B., Xu, L., 2012. Occurrence of sulfonamide and tetracycline-resistant bacteria and resistance genes in aquaculture environment. *Water Res.* 46, 2355–2364.
- Germond, A., Kim, S.J., 2015. Genetic diversity of Oxytetracycline resistant bacteria and tet(M) genes in two major coastal areas of South Korea. *J. Glob. Antimicrob. Resist.* 3, 166–173.
- Giang, C.N.D., Sebesvari, Z., Renaud, F., Rosendahl, I., Minh, Q.H., Amelung, W., 2015. Occurrence and dissipation of the antibiotics sulfamethoxazole, sulfadiazine, trimethoprim, and enrofloxacin in the Mekong Delta, Vietnam. *PLoS One* 10, e0131855.
- Granada, L., Sousa, N., Lopes, S., Lemos, M.F.L., 2016. Is integrated multitrophic aquaculture the solution to the sectors' major challenges? – A review. *Rev. Aquac.* 8, 283–300.
- Gros, M., Marti, E., Balcázar, J.L., Boy-Roura, M., Busquets, A., Colón, J., Sánchez-Melsió, A., Lekunberri, I., Borrego, C.M., Ponsá, S., Petrovic, M., 2019. Fate of pharmaceuticals and antibiotic resistance genes in a full-scale on-farm livestock waste treatment plant. *J. Hazard. Mater.* 378, 120716 <https://doi.org/10.1016/j.jhazmat.2019.05.109>.

- Hassan, M.N., Rahman, M., Hossain, M.B., Hossain, M.M., Mendes, R., Nowsad, A., 2013. Monitoring the presence of Chloramphenicol and nitrofurans metabolites in cultured prawn, shrimp and feed in the Southwest coastal region of Bangladesh. *Egypt. J. Aquat. Res.* 39, 51–58.
- Hawkey, P., 2008. The growing burden of antimicrobial resistance. *J. Antimicrob. Chemother.* 62, i1–i9.
- He, S., Zhou, Z., Liu, Y., Cao, Y., Meng, K., Shi, P., et al., 2012a. Dietary betaine and the antibiotic florfenicol influence the intestinal autochthonous bacterial community in hybrid tilapia (*Oreochromis niloticus* ♀ × *O. aureus* ♂)? *World J. Microbiol. Biotechnol.* 28, 785–791.
- He, Y., Tang, Y., Sun, F., Chen, L., 2015. Detection and characterization of integrative and conjugative elements (ICEs)-positive *Vibrio cholerae* isolates from aqua cultured shrimp and the environment in Shanghai, China. *Mar. Pollut. Bull.* 101, 526–532.
- Henríquez-Núñez, H., Evrard, O., Kronvall, G., Avendaño-Herrera, R., 2012. Antimicrobial susceptibility and plasmid profiles of *Flavobacterium psychrophilum* strains isolated in Chile. *Aquaculture* 354–355, 38–44.
- Heuer, O.E., Kruse, H., Grave, K., Collignon, P., Karunasagar, I., Angulo, F.J., 2009. Human health consequences of use of antimicrobial agents in aquaculture. *Clin. Infect. Dis.* 49, 1248–1253.
- Hong, B., Ba, Y., Niu, L., Lou, F., Zhang, Z., Liu, H., et al., 2018. A comprehensive research on antibiotic resistance genes in microbiota of aquatic animals. *Front. Microbiol.* 9, 1617.
- Hossain, A., Nakamichi, S., Habibullah-Al-Mamun, M., Tani, K., Masunaga, S., Matsuda, H., 2017. Occurrence, distribution, ecological and resistance risks of antibiotics in surface water of finfish and shellfish aquaculture in Bangladesh. *Chemosphere* 188, 329–336.
- Huang, S., Wang, L., Liu, L., Hou, Y., Li, L., 2015. Nanotechnology in agriculture, livestock, and aquaculture in China. A review. *Agron. Sustain. Dev.* 35, 369–400.
- Ishida, Y., Ahmed, A.M., Mahfouz, N.B., Kimura, T., El-Khodery, S.A., Moawad, A.A., et al., 2010. Molecular analysis of antimicrobial resistance in gram-negative bacteria isolated from fish farms in Egypt. *J. Vet. Med. Sci.* 72, 727–734.
- Jang, H.M., Kim, Y.B., Choi, S., Lee, Y., Shin, S.G., Unno, T., et al., 2018. Prevalence of antibiotic resistance genes from effluent of coastal aquaculture, South Korea. *Environ. Pollut.* 233, 1049–1057.
- Justino, C.L.L., Duarte, K.R., Freitas, A.C., Panteleitchouk, T.S.L., Duarte, A.C., Rocha-Santos, T.A.P., 2016. Contaminants in aquaculture: overview of analytical techniques for their determination. *Trends Anal. Chem.* 80, 293–310.
- Kim, Y.B., Jeon, J.H., Choi, S., Shin, J., Lee, Y., Kim, Y.M., 2018. Use of a filtering process to remove solid waste and antibiotic resistance genes from effluent of a flow-through fish farm. *Sci. Total Environ.* 615, 289–296.
- Lafferty, K.D., Harvell, C.D., Conrad, J.M., Friedman, C.S., Kent, M.L., Kuris, A.M., et al., 2015. Infectious diseases affect marine fisheries and aquaculture economics. *Ann. Rev. Mar. Sci.* 7, 471–496.
- Lai, H.T., Lin, J.J., 2009. Degradation of oxolinic acid and flumequine in aquaculture pond waters and sediments. *Chemosphere* 75, 462–468.
- Lalumera, G.M., Calamari, D., Galli, P., Castiglioni, S., Crosa, G., Fanelli, R., 2004. Preliminary investigation on the environmental occurrence and effects of antibiotics used in aquaculture in Italy. *Chemosphere* 54, 661–668.
- Le, T.X., Munekage, Y., 2004. Residues of selected antibiotics in water and mud from shrimp ponds in mangrove areas in Vietnam. *Mar. Pollut. Bull.* 49, 922–929.
- Letchumanan, V., Pusparajah, P., Tan, L.T., Yin, W.F., Lee, L.H., Chan, K.G., 2015a. Occurrence and antibiotic resistance of *Vibrio parahaemolyticus* from Shellfish in Selangor, Malaysia. *Front. Microbiol.* 6, 1417.
- Letchumanan, V., Yin, W.F., Lee, L.H., Chan, K.G., 2015b. Prevalence and antimicrobial susceptibility of *Vibrio parahaemolyticus* isolated from retail shrimps in Malaysia. *Front. Microbiol.* 6, 33.
- Li, H., Duan, M., Gu, J., Zhang, Y., Qian, X., Ma, J., Zhang, R., Wang, X., 2017. Effects of bamboo charcoal on antibiotic resistance genes during chicken manure composting. *Ecotoxicol. Environ. Saf.* 140, 1–6. <https://doi.org/10.1016/j.ecoenv.2017.01.007>.
- Li, Z., Hu, Y., Yang, Y., Lu, Z., Wang, Y., 2018. Antimicrobial resistance in livestock: Antimicrobial peptides provide a new solution for a growing challenge. *Anim. Front. Rev. Mag. Anim. Agric.* 8 (2), 21–29. <https://doi.org/10.1093/af/vfy005>.
- Li, Juan-Ying, Wen, Ju, Chen, Y., Wang, Q., Yin, J., 2021. Antibiotics in cultured freshwater products in Eastern China: Occurrence, human health risks, sources, and bioaccumulation potential. *Chemosphere* 264, 128441.
- Li, Z.J., 2008. Advantages and disadvantages and strategies of antibiotic application in aquaculture. *Guizhou Anim. Sci. Vet. Med.* 32, 23–24.
- Liang, X.M., Nie, X.P., Shi, Z., 2013. Preliminary studies on the occurrence of antibiotic resistance genes in typical aquaculture area of the Pearl River Estuary. *Environ. Sci.* 34, 4073–4080.
- Liao, H.P., Lu, X.M., Rensing, C., Friman, V.P., Geisen, S., Chen, Z., Yu, Z., Wei, Z., Zhou, S.G., Zhu, Y.G., 2018. Hyperthermophilic composting accelerates the removal of antibiotic resistance genes and mobile genetic elements in sewage sludge. *Environ. Sci. Technol.* 52 (1), 266–276. <https://doi.org/10.1021/acs.est.7b04483>.
- Liu, X., Steele, J.C., Meng, X.Z., 2017. Usage, residue, and human health risk of antibiotics in Chinese aquaculture: a review. *Environ. Pollut.* 223, 161–169.
- Ma, J., Wang, S., Wang, P., Ma, L., Chena, X., Xu, R., 2006. Toxicity assessment of 40 herbicides to the green alga *Raphidocelis subcapitata*. *Ecotoxicol. Environ. Saf.* 63, 456–462.
- Marshall, B.M., Levy, S.B., 2011. Food animals and antimicrobials: impacts on human health. *Clin. Microbiol. Rev.* 24, 718–733.
- Marti, E., Huerta, B., Rodríguez-Mozaz, S., Barcelo, D., Marce, R., Balcazar, J.L., 2018. Abundance of antibiotic resistance genes and bacterial community composition in wild freshwater fish species. *Chemosphere* 196, 115–119.
- Martínez, J.L., 2009. The role of natural environments in the evolution of resistance traits in pathogenic bacteria. *Proc. R. Soc. B Biol. Sci.* 276, 2521–2530.
- Martínez-Porchas, M., Martínez-Cordova, L.R., 2012. World aquaculture: environmental impacts and troubleshooting alternatives. *Sci. World J.* 2012, 1–9.
- Midtlyng, P.J., Grave, K., Horsberg, T.E., 2011. What has been done to minimize the use of antibacterial and antiparasitic drugs in Norwegian aquaculture? *Aquac. Res.* 42, 28–34.
- Millanao, A.R., Barrientos-Schaffeld, C., Siegel-Tike, C.D., Tomova, A., Ivanova, L., Godfrey, H.P., et al., 2018. Antimicrobial resistance in Chile and the one health paradigm: dealing with threats to human and veterinary health resulting from antimicrobial use in Salmon aquaculture and the clinic. *Rev. Chil. Infectol.* 35, 299–303.
- Mishra, R.K., Verma, D.K., Ravindra Yadav, M.K., Pradhan, P.K., Swaminathan, T.R., Sood, N., 2017. Bacterial diversity and antibiotic resistance in a wetland of Lakhimpur-Kheri, Uttar Pradesh, India. *J. Environ. Biol.* 38, 55–66.
- Molnar, J.L., Gamboa, R.L., Revenga, C., Spalding, M.D., 2008. Assessing the global threat of invasive species to marine biodiversity. *Front. Ecol. Environ.* 6, 485–492.
- Monteiro, S.H., Garcia, F., Gozi, K.S., Romera, D.M., Francisco, J.G., Moura-Andrade, G. C.R., et al., 2016. Relationship between antibiotic residues and occurrence of resistant bacteria in Nile tilapia (*Oreochromis niloticus*) cultured in cage-farm. *J. Environ. Sci. Health Part B-Pestic. Food Contam. Agric. Wastes* 51, 817–823.
- Moreau, D.T., Neis, B., 2009. Occupational health and safety hazards in Atlantic Canadian aquaculture: laying the groundwork for prevention. *Mar. Policy* 33, 401–411.
- Murray, A.G., 2013. Epidemiology of the spread of viral diseases under aquaculture. *Curr. Opin. Virol.* 3, 74–78.
- Nguyen, H.N.K., Van, T.T.H., Nguyen, H.T., Smooker, P.M., Shimeta, J., Coloe, P.J., 2014. Molecular characterization of antibiotic resistance in *Pseudomonas* and *Aeromonas* isolates from catfish of the Mekong Delta, Vietnam. *Vet. Microbiol.* 171, 397–405.
- Noorli, A., Ghazali, F.M., Cheah, Y.K., Tuan Zainazor, T.C., Wong, W.C., Tunung, R., et al., 2011. Antibiotic resistance and biosafety of *Vibrio cholerae* and *Vibrio parahaemolyticus* from freshwater fish at retail level. *Int. Food Res. J.* 18, 1523–1530.
- Novo, A., Manaia, C.M., 2010. Factors influencing antibiotic resistance burden in municipal wastewater treatment plants. *Appl. Microbiol. Biotechnol.* 87 (3), 1157–1166. <https://doi.org/10.1007/s00253-010-2583-6>.
- OECD, 2011. Guidelines for the Testing of Chemicals, Section 2. Test No. 201: Freshwater Alga and Cyanobacteria, Growth Inhibition Test. PDF Edition (ISSN:2074-5761), p. 25.
- Okeke, E.S., Chukwudozie, K.I., Nyaruaba, R., et al., 2022. Antibiotic resistance in aquaculture and aquatic organisms: a review of current nanotechnology applications for sustainable management. *Environ. Sci. Pollut. Res.* 29, 69241–69274.
- Osman, K.M., Ali, M.N., Radwan, I., El Hofy, F., Abed, A.H., Orabi, A., et al., 2016. Dispersion of the vancomycin resistance genes van A and van C of *Enterococcus* isolated from Nile Tilapia on retail sale: a public health hazard. *Front. Microbiol.* 7, 1354.
- Osman, K.M., Al-Maary, K.S., Mubarak, A.S., Dawoud, T.M., Moussa, M.I., Ibrahim, M.D. S., et al., 2017. Characterization and susceptibility of streptococci and enterococci isolated from Niletilapia (*Oreochromis niloticus*) showing septicemia in aquaculture and wild sites in Egypt. *BMC Vet. Res.* 13, 357.
- Páez-Osuna, F., 2001. The environmental impact of shrimp aquaculture: causes, effects, and mitigating alternatives. *Environ. Manage.* 28, 131–140.
- Park, K., Kwak, I.S., 2018. Disrupting effects of antibiotic sulfathiazole on developmental process during sensitive life-cycle stage of *Chironomus riparius*. *Chemosphere* 190, 25–34.
- Peng, S., Li, H., Song, D., Lin, X., Wang, Y., 2018. Influence of zeolite and superphosphate as additives on antibiotic resistance genes and bacterial communities during factory-scale chicken manure composting. *Bioresour. Technol.* 263, 393–401. <https://doi.org/10.1016/j.biortech.2018.04.107>.
- Pham, T.T.H., Rossi, P., Dinh, H.D.K., Pham, N.T.A., Tran, P.A., Ho, T., et al., 2018. Analysis of antibiotic multi-resistant bacteria and resistance genes in the effluent of an intensive shrimp farm (Long An, Vietnam). *J. Environ. Manage.* 214, 149–156.
- Phu, T.M., Phuong, N.T., Dung, T.T., Hai, D.M., Son, V.N., Rico, A., et al., 2016. An evaluation of fish health-management practices and occupational health hazards associated with Pangasius catfish (*Pangasius hypophthalmus*) aquaculture in the Mekong Delta, Vietnam. *Aquac. Res.* 47, 2778–2794.
- Rahiman, K.M.M., Hatha, A.A.M., Selvam, A.D.G., Thomas, A.P., 2016. Relative prevalence of antibiotic resistance among heterotrophic bacteria from natural and culture environments offreshwater Prawn, *Macrobrachium rosenbergii*. *J. World Aquac. Soc.* 47, 470–480.
- Rahman, M., Huys, G., Kühn, I., Rahman, M., Möllby, R., 2009. Prevalence and transmission of antimicrobial resistance among *Aeromonas* populations from a duckweed aquaculture based hospital sewage water recycling system in Bangladesh. *Antonie van Leeuwenhoek. Int. J. Gen. Mol. Microbiol.* 96, 313–321.
- Rajitha, K., Mukherjee, C.K., Vinu Chandran, R., 2007. Applications of remote sensing and GIS for sustainable management of shrimp culture in India. *Aquacult. Eng.* 36, 1–17.
- Reboucas, R.H., de Sousa, O.V., Lima, A.S., Vasconcelos, F.R., de Carvalho, P.B., Vieira, R., 2011. Antimicrobial resistance profile of *Vibrio* species isolated from marine shrimp farming environments (*Litopenaeus vannamei*) at Ceara, Brazil. *Environ. Res.* 111, 21–24.
- Reungprach, H., Kesomchandra, J., 1983. *In vitro* drug susceptibility studies of bacterial isolated during the epizootic in 1982-1983 to some antibiotics and sulfas. *Thai Fish. Gazette* 36, 264–267.
- Rico, A., Phu, T.M., Satapornvanit, K., Min, J., Shahabuddin, A.M., Henriksson, P.J.G., et al., 2013. Use of veterinary medicines, feed additives and probiotics in four major internationally traded aquaculture species farmed in Asia. *Aquaculture* 412, 231–243.

- Rocha, R.S., Sousa, O.V., Vieira, R., 2016. Multidrug-resistant *Vibrio* associated with an estuary affected by shrimp farming in Northeastern Brazil. *Mar. Pollut. Bull.* 105, 337–340.
- Saitanu, K., Chalarak, C., 1983. Ulcer disease in catfish (*Clarias batrachus*): a therapeutic study. *J. Aquat. Anim. Dis.* 6, 9–17.
- Saitanu, K., Wongsawang, S., 1982. Red-sore disease in carp (*Cyprinus carpio*). *J. Aquat. Anim. Dis.* 5, 79–86.
- Samuelsen, O.B., Lunestad, B.T., Farestveit, E., Grefsrud, E.S., Hannisdal, R., Holmelid, B., et al., 2014. Mortality and deformities in European lobster (*Homarus gammarus*) juveniles exposed to the antiparasitic drug teflubenzuron. *Aquat. Toxicol.* 149, 8–15.
- Santos, L., Araújo, A.N., Fachini, A., Pena, A., Delerue-Matos, C., Montenegro, M., 2010. Ecotoxicological aspects related to the presence of pharmaceuticals in the aquatic environment. *J. Hazard. Mater.* 175, 45–95.
- Sarmah, A.K., Meyer, M.T., Boxall, A.B.A., 2006. A global perspective on the use, sales, exposure pathways, occurrence, fate and effects of veterinary antibiotics (VAs) in the environment. *Chemosphere* 65, 725–759.
- Schwarz, S., Cloeckaert, A., Roberts, M.C., 2006. Mechanisms and Spread of Bacterial Resistance to Antimicrobial Agents. *Antimicrobial Resistance in Bacteria of Animal Origin*. American Society of Microbiology, pp. 73–98.
- Selvam, A., Wong, J.W.C., 2017. 12 – Degradation of antibiotics in livestock manure during composting. *J. W.-C. Wong, R. D. Tyagi, & A. Pandey Current Developments in Biotechnology & Bioengineering*. Elsevier, pp. 267–292.
- Shah, S.Q., Cabello, F.C., L'Abée-Lund, T.M., Tomova, A., Godfrey, H.P., Buschmann, A. H., et al., 2014. Antimicrobial resistance and antimicrobial resistance genes in marine bacteria from salmon aquaculture and non-aquaculture sites. *Environ. Microbiol.* 16, 1310–1320.
- Shao, Y., Wang, Y., Yuan, Y., Xie, Y., 2021. A systematic review on antibiotics misuse in livestock and aquaculture and regulation implications in China. *Sci. Total Environ.* 798, 149205.
- Shimizu, A., Takada, H., Koike, T., Takeshita, A., Saha, M., Nakada, N., 2013. Ubiquitous occurrence of sulfonamides in tropical Asian waters. *Sci. Total Environ.* 452, 108–115.
- Sneha, K.G., Anas, A., Jayalakshmy, K.V., Jasmin, C., Das, P.V.V., Pai, S.S., 2016. Distribution of multiple antibiotic resistant *Vibrio* spp. across Palk Bay. *Reg. Stud. Mar. Sci.* 3, 242–250.
- Son, T.T.D., Petersen, A., Truong, D.V., Huong, T.T.C., Dalsgaard, A., 2011. Impact of medicated feed on the development of antimicrobial resistance in bacteria at integrated pig-fish farms in Vietnam. *Appl. Environ. Microbiol.* 77, 4494–4498.
- Song, C., Zhang, C., Fan, L., Qiu, L., Wu, W., Meng, S., et al., 2016. Occurrence of antibiotics and their impacts to primary productivity in fishponds around Tai Lake, China. *Chemosphere* 161, 127–135.
- Stratev, D., Odeyemi, O.A., 2016. Antimicrobial resistance of *Aeromonas hydrophila* isolated from different food sources: a mini review. *J. Infect. Public Health* 9, 535–544.
- Su, H.C., Liu, S., Hu, X.J., Xu, X.R., Xu, W.J., Xu, Y., et al., 2017. Occurrence and temporal variation of antibiotic resistance genes (ARGs) in shrimp aquaculture: ARGs dissemination from farming source to reared organisms. *Sci. Total Environ.* 607, 357–366.
- Sun, Y., Liu, C.S., Sun, L., 2010. Isolation and analysis of the vaccine potential of an attenuated *Edward siellatarda* strain. *Vaccine* 28, 6344–6350.
- Tamminen, M., Karkman, A., Lohmus, A., Muziasari, W.I., Takasu, H., W.S., et al., 2011. Tetracycline resistance genes persist at aquaculture farms in the absence of selection pressure. *Environ. Sci. Technol.* 45, 386–391.
- Tang, K.L., Caffrey, N.P., Nobrega, D.B., Cork, S.C., Ronksley, P.E., Barkema, H.W., Polachek, A.J., Ganshorn, H., Sharma, N., Kellner, J.D., Ghali, W.A., 2017. Restricting the use of antibiotics in food-producing animals and its associations with antibiotic resistance in food-producing animals and human beings: A systematic review and meta-analysis. *Lancet Planet Health* 1 (8), e316–e327. [https://doi.org/10.1016/S2542-5196\(17\)30141-9](https://doi.org/10.1016/S2542-5196(17)30141-9).
- Tello, A., Corner, R.A., Telfer, T.C., 2010. How do land-based salmonid farms affect stream ecology? *Environ. Pollut.* 158, 1147–1158.
- Tendencia, E.A., De la Peña, L.D., 2001. Antibiotic resistance of bacteria from shrimp ponds. *Aquaculture* 195, 193–204.
- Thai, P.K., Ky, L.X., Binh, V.N., Nhung, P.H., Nhan, P.T., Hieu, N.Q., et al., 2018. Occurrence of antibiotic residues and antibiotic-resistant bacteria in effluents of pharmaceutical manufacturers and other sources around Hanoi, Vietnam. *Sci. Total Environ.* 645, 393–400.
- Thuy, H.T.T., Nguyen, T.D., 2013. The potential environmental risks of pharmaceuticals in Vietnamese aquatic systems: case study of antibiotics and synthetic hormones. *Environ. Sci. Pollut. Res.* 20, 8132–8140.
- Tomova, A., Ivanova, L., Buschmann, A.H., Rioseco, M.L., Kalsi, R.K., Godfrey, H.P., et al., 2015. Antimicrobial resistance genes in marine bacteria and human uropathogenic *Escherichia coli* from a region of intensive aquaculture. *Environ. Microbiol. Rep.* 7, 803–809.
- Tušečljak, N., Dutil, L., Rajić, A., Uhlund, F., McClure, C., St-Hilaire, S., 2013. Antimicrobial use and resistance in aquaculture: findings of a globally administered survey of aquaculture-allied professionals. *Zoonoses Public Health* 60, 426–436.
- Wang, Q., Cheng, L., Liu, J., Li, Z., Xie, S., De Silva, S.S., 2015. Freshwater aquaculture in PR China: trends and prospects. *Rev. Aquac.* 7, 283–302.
- White, D.G., McDermott, P.F., 2009. Antimicrobial resistance in food-borne pathogens. *Food-Borne Microbes*. American Society of Microbiology, Washington, DC, pp. 231–265.
- Wollenberger, L., Halling-Sørensen, B., Kusk, K.O., 2000. Acute and chronic toxicity of veterinary antibiotics to *Daphnia magna*. *Chemosphere* 40, 723–730.
- Xu, L., Zhang, H., Xiong, P., Zhu, Q., Liao, C., Jiang, G., 2021. Occurrence, fate, and risk assessment of typical tetracycline antibiotics in the aquatic environment: A review. *Sci. Total Environ.* 753, 141975.
- Yasser, E.L.N., Adli, A., 2015. Toxicity of single and mixtures of antibiotics to Cyanobacteria. *J. Environ. Anal. Toxicol.* 5, 274–281.
- Yi, S.W., Chung, T.H., Joh, S.J., Park, C., Park, B.Y., Shin, G.W., 2014. High prevalence of Bla CTX-M group genes in *Aeromonas dhakensis* isolated from aquaculture fish species in South Korea. *J. Vet. Med. Sci.* 76, 1589–1593.
- Yi, Z., Yang, Q.E., Zhou, X., Wang, F.H., Muurinen, J., Virta, M.P., Brandt, K.K., Zhu, Y.G., 2020. Antibiotic resistome in the livestock and aquaculture industries: Status and solutions. *Crit. Rev. Environ. Sci. Technol.* <https://doi.org/10.1080/10643389.2020.1777815>.
- Zhang, Q., Jia, A., Wan, Y., Liu, H., Wang, K., Peng, H., et al., 2014. Occurrences of three classes of antibiotics in a natural river basin: association with antibiotic-resistant *Escherichia coli*. *Environ. Sci. Technol.* 48, 14317–14325.
- Zhang, J., Lu, T., Shen, P., Sui, Q., Zhong, H., Liu, J., Tong, J., Wei, Y., 2019. The role of substrate types and substrate microbial community on the fate of antibiotic resistance genes during anaerobic digestion. *Chemosphere* 229, 461–470. <https://doi.org/10.1016/j.chemosphere.2019.05.036>.
- Zhang, J., Zhang, X., Zhou, Y., Han, Q., Wang, X., Song, C., Wang, S., Zhao, S., 2023. Occurrence, distribution and risk assessment of antibiotics at various aquaculture stages in typical aquaculture areas surrounding the Yellow Sea. *J. Environ. Sci.* 126, 621–632.
- Zheng, B., Su, S.X., 2010. Status, effects and strategies of antibiotic residues Chinese. *J. Anal. Lab.* 29, 285–287.
- Zounková, R., Klimešová, Z., Nepechalová, L., Hilscherová, B.L., 2011. Complex evaluation of ecotoxicity and genotoxicity of antimicrobials oxytetracycline and flumequine used in aquaculture. *Environ. Toxicol. Chem.* 30, 1184–1189.
- Zulkifli, Y., Alitheen, N.B., Raha, A.R., Yeap, S.K., Marlina, S.R., Nishibuchi, M., 2009. Antibiotic resistance and plasmid profiling of *Vibrio parahaemolyticus* isolated from cockles in Padang, Indonesia. *Int. Food Res. J.* 16, 53–58.